

# **Implementing an Ambient Phytoplankton Monitoring Program in Puget Sound Central Basin**

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# Marine Ambient Program

- ◆ King County Marine and Sediment Assessment Group
- ◆ Long-term marine monitoring program - since 1995
- ◆ Water quality in the Central Puget Sound Basin
- ◆ Physical, chemical and biological parameters - monthly
- ◆ Marine Phytoplankton monitoring    ***NEW!! 2008***

# Goals of Marine Phytoplankton Monitoring

**Measure/predict impact of climate or regional stressors on community structure**

- ◆ Assess relative abundance of major phytoplankton taxa during the bloom season
- ◆ Document the timing of seasonal shifts in major taxa
- ◆ Investigate relationships between physical/chemical parameters and species relative abundance
- ◆ Detect long-term changes in community composition

# More Goals !

- ◆ Create an electronic image library of Puget Sound phytoplankton species
- ◆ Detect occurrence and environmental conditions that lead to blooms of HAB species in Puget Sound
- ◆ Expand the program and build extensive database

# Sample Collection

## 3 Locations in the Central Basin

### Point Jefferson

Long-term station

Open, North

Sample 2 depths

### East Passage

Long-term station

Open, South

Sample 2 depths

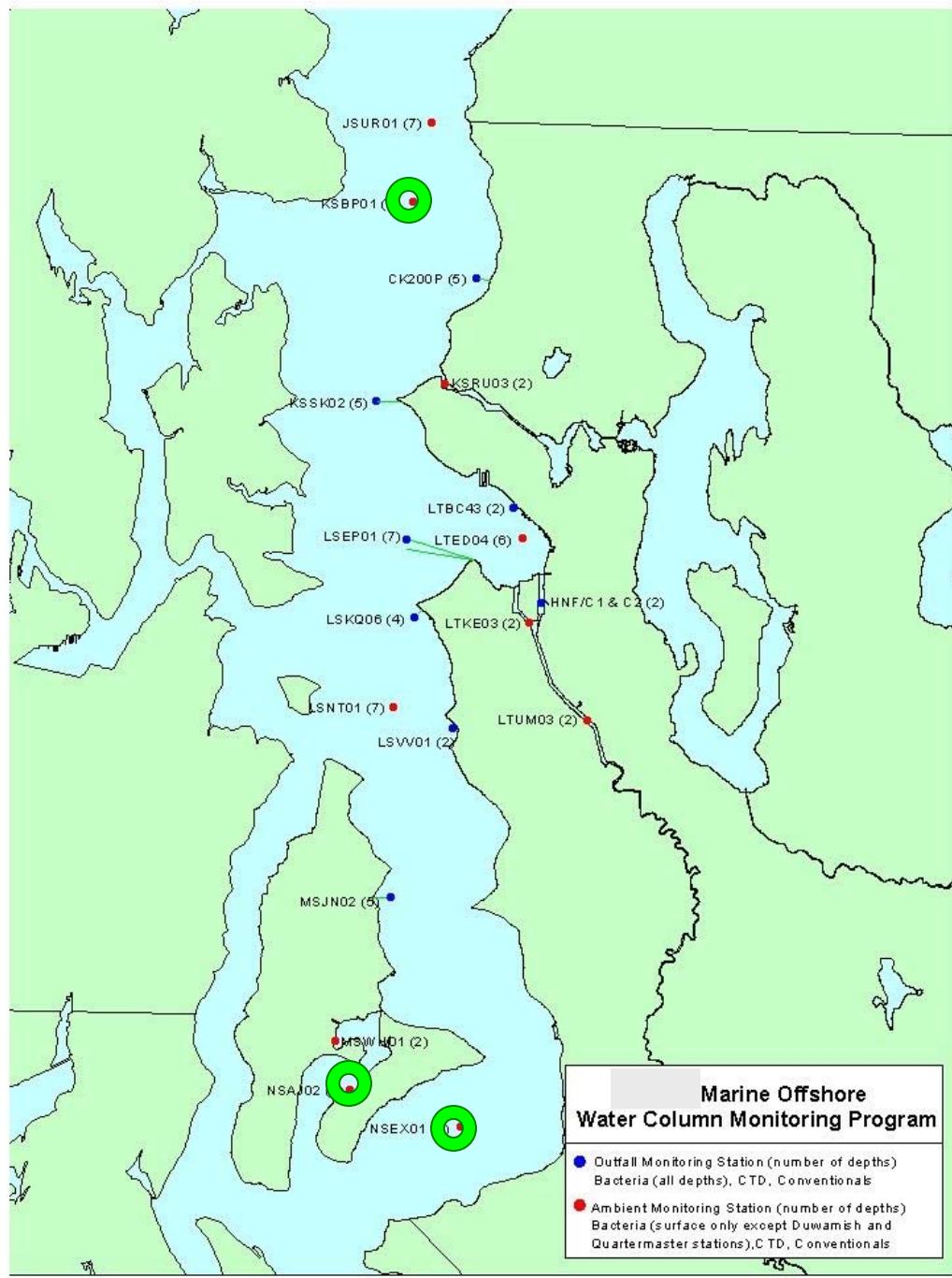
### Quartermaster Harbor

Shallow protected  
embayment with  
poor flushing

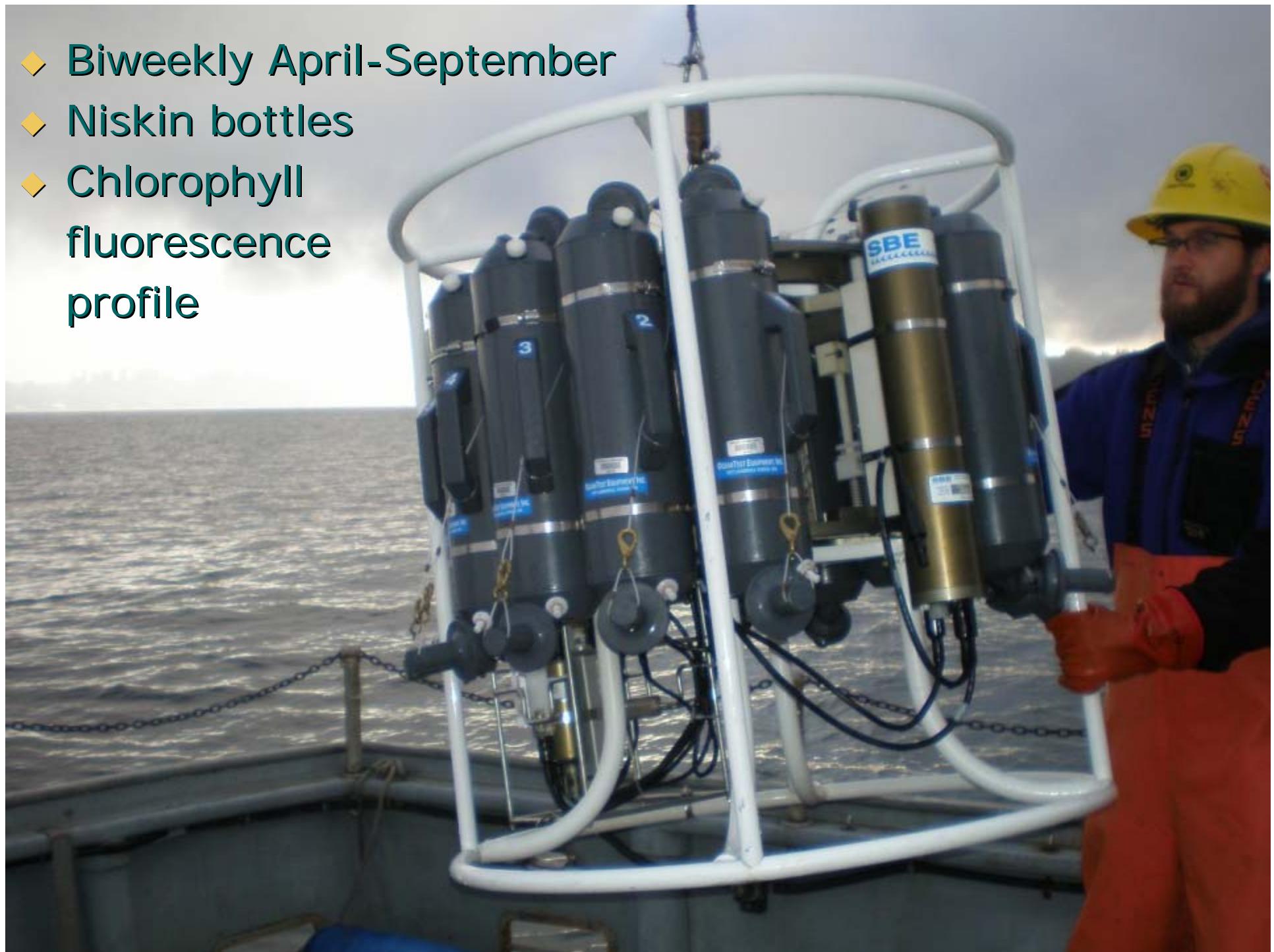
Sample 1 depth

Telemetered  
mooring





- ◆ Biweekly April-September
- ◆ Niskin bottles
- ◆ Chlorophyll fluorescence profile





# Laboratory Methods

## 1. List of taxa

Species

Genus

Larger category



## 2. Semi-quantitative evaluation at genus level - based on cell #

Present <25%

Subdominant 25-50%

Dominant >50%



# Live Samples

- ◆ Used for species identification
- ◆ Concentration by reverse filtration  
1 or 5 µm mesh      ~50x



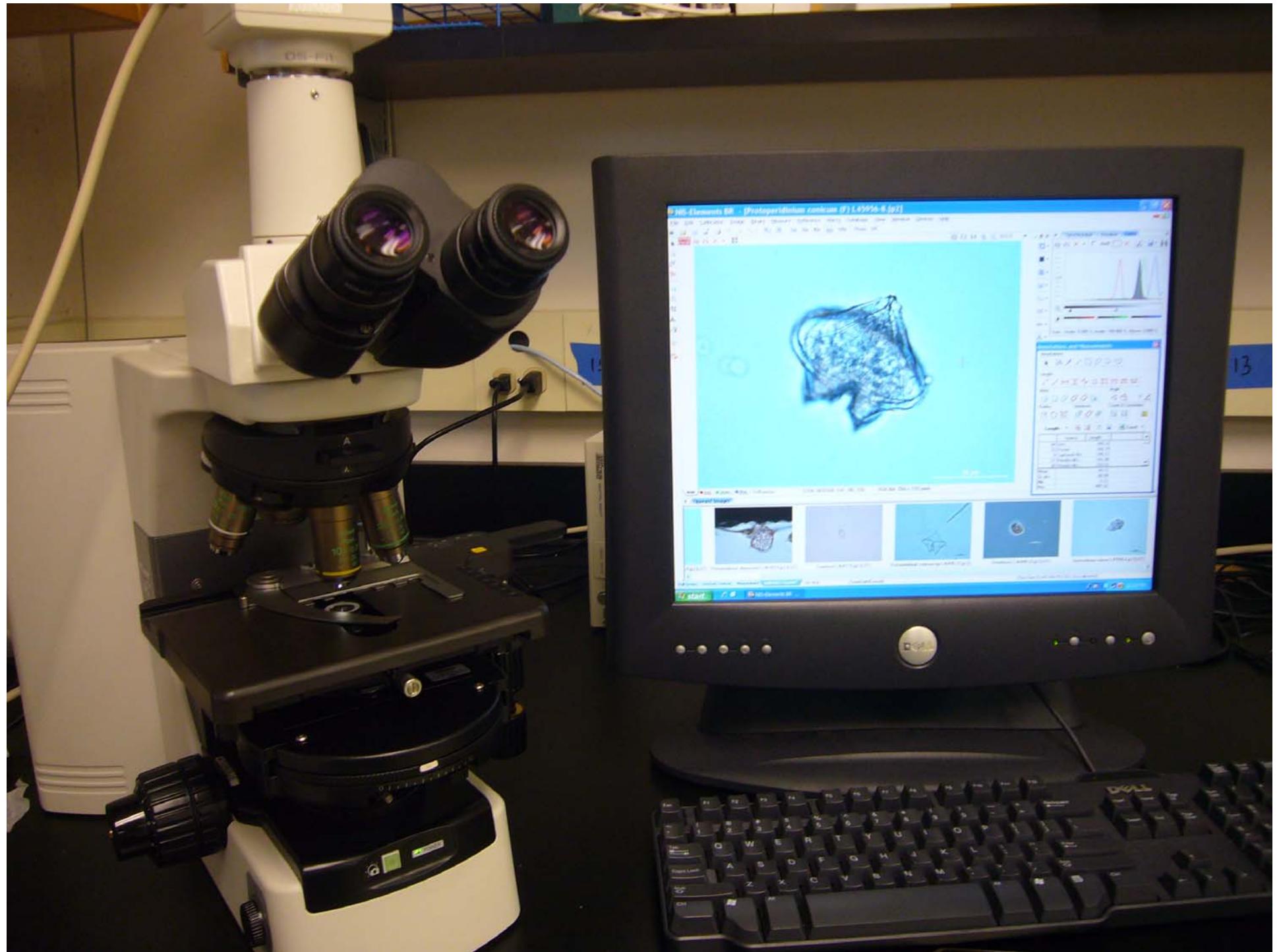
# Preserved Samples

For archiving and semi-quantitative analysis



Settling in Formalin





# Nanoplankton Counting Chamber





Photograph: K. Newell

The author has a Ph.D. in marine botany from the University of Washington and learned phytoplankton as a Fulbright student at the University of Oslo, Norway. For the last four decades, she has studied marine phytoplankton primarily in the Arctic and North Pacific oceans. Dr Horner has been associated with the School of Oceanography at the University of Washington since 1974 where she has worked on a variety of projects ranging from the biology of polar phytoplankton and ice algae to the taxonomy and ecology of harmful algal blooms.

Dr Horner is a member of the Phycological Society of America, the British Phycological Society, the International Phycological Society, the International Society for Diatom Research, the International Society for the Study of Harmful Algae, and is a Fellow of the Arctic Institute of North America.

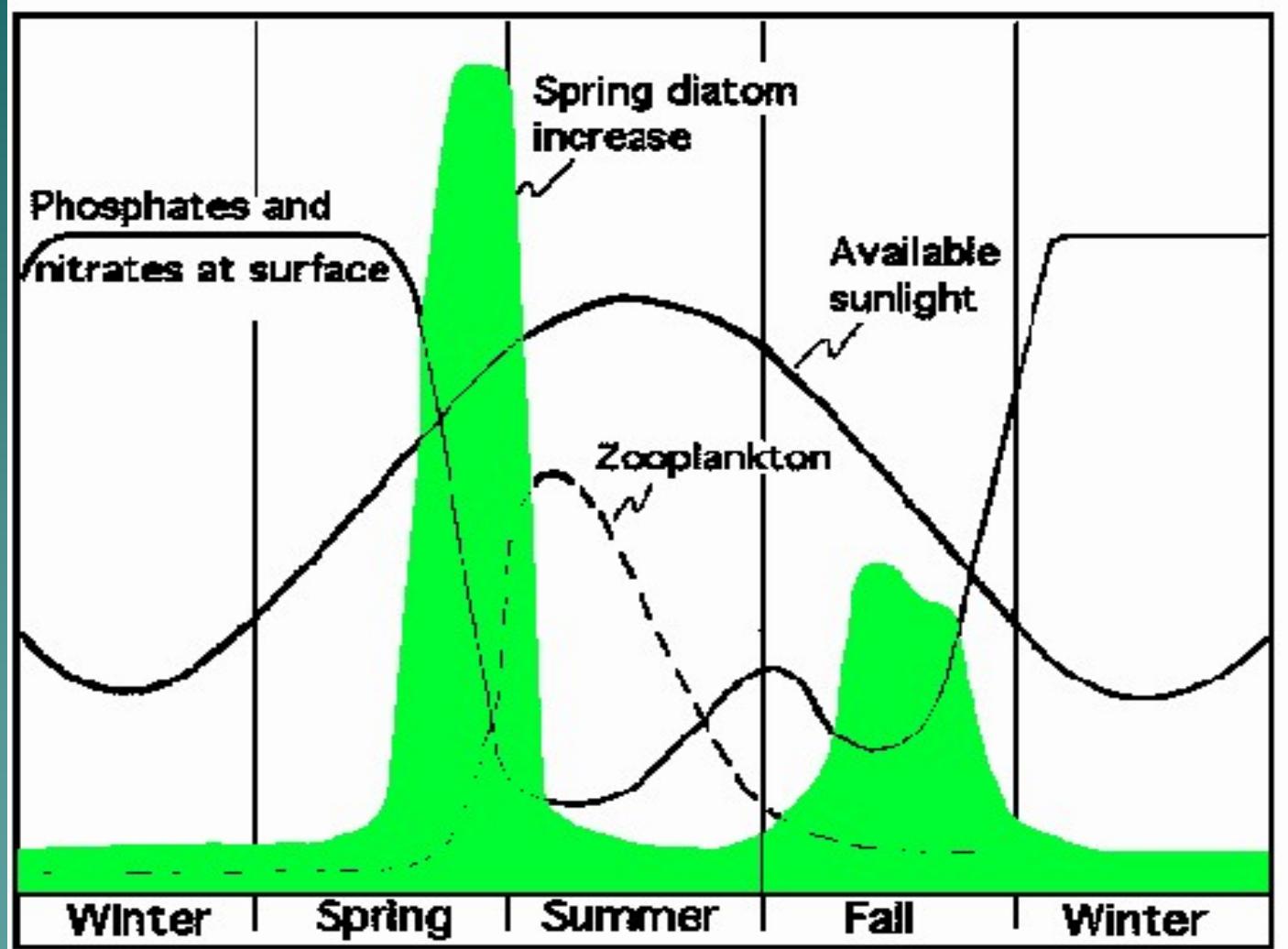
## A TAXONOMIC GUIDE TO SOME COMMON MARINE PHYTOPLANKTON

Rita A. Horner



Biopress Limited

# Typical Seasonal Pattern



# Some Results/Observations

- ◆ Number of species
- ◆ Dominant taxa
- ◆ Harmful species?

# List of taxa in LIMS

<b>DIATOMS-CENTRIC</b>	<b>DIATOMS-PENNATE</b>	<b>CHRYSTOPHYTES</b>
<i>Actinoptychus senarius</i>	<i>Asterionellopsis glacialis</i>	<i>Meringosphaera mediterranea</i>
<i>Asteromphalus heptactis</i>	<i>Cylindrotheca closterium</i>	
<i>Aulacodiscus kittonii</i>	<i>Navicula</i> sp.	
<i>Cerataulina pelagica</i>	<i>Nitzschia acicularis</i>	
<i>Chaetoceros (Hyalochaete) sp.</i>	<i>Pleurosigma</i> sp.	<b>EBRIDEAN</b>
<i>Chaetoceros (Phaeocerus) sp.</i>	<i>Pseudo-nitzschia</i> sp. (large)	<i>Ebria tripartita</i>
<i>Chaetoceros affinis</i>	<i>Pseudo-nitzschia</i> sp. (small)	
<i>Chaetoceros concavicornis</i>	<i>Thalassionema nitzschiooides</i>	
<i>Chaetoceros convolutus</i>	<i>Tropidoneis antarctica</i>	<b>PRYMNESIOPHYTES</b>
<i>Chaetoceros curvisetus</i>	unidentified pennate	<i>Phaeocystis</i> sp.
<i>Chaetoceros danicus</i>		
<i>Chaetoceros debilis</i>		
<i>Chaetoceros decipiens</i>	<b>DINOFLAGELLATES</b>	
<i>Chaetoceros diadema</i>	<i>Akashiwo sanguinea</i>	
<i>Chaetoceros didymus</i>	<i>Alexandrium catenula</i>	
<i>Chaetoceros laciniosus</i>	<i>Alexandrium</i> sp.	
<i>Chaetoceros lorenzianus</i>	<i>Amylax triacantha</i>	
<i>Chaetoceros radicans</i>	<i>Ceratium fusus</i>	
<i>Chaetoceros similis</i>	<i>Ceratium</i> sp.	
<i>Chaetoceros socialis</i>	<i>Dinophysis acuminata</i>	
<i>Chaetoceros teres</i>	<i>Dinophysis acuta/norvegica</i>	
<i>Chaetoceros vanheurckii</i>	<i>Dinophysis fortii</i>	
<i>Corethron hystrix</i>	<i>Dinophysis rotundata</i>	
<i>Coscinodiscus centralis</i>	<i>Dinophysis</i> sp.	
<i>Coscinodiscus concinnus</i>	<i>Dissodinium pseudolunula</i>	
<i>Coscinodiscus curvatus</i>	<i>Gonyaulax</i> sp.	
<i>Coscinodiscus granii</i>	<i>Gymnodinium</i> sp.	
<i>Coscinodiscus marginatus</i>	<i>Gyrodinium</i> sp.	
<i>Coscinodiscus oculus-iridis</i>	<i>Gyrodinium spirale</i>	
<i>Coscinodiscus sp.</i>	<i>Heterocapsa triquetra</i>	
<i>Coscinodiscus wailesii</i>	<i>Minuscula bipes</i>	
<i>Dactyliosolen fragilissimus</i>	<i>Nematodinium armatum</i>	
<i>Detonula pumila</i>	<i>Noctiluca scintillans</i>	
<i>Ditylum brightwellii</i>	<i>Oxyphyasis oxytoxoides</i>	
<i>Eucampia zodiacus</i>	<i>Polykrikos schwartzii</i>	
<i>Guinardia delicatula</i>	<i>Prorocentrum gracile</i>	
<i>Guinardia striata</i>	<i>Prorocentrum micans</i>	
<i>Hemiaulus hauckii</i>	<i>Protoceratium reticulatum</i>	
<i>Lauderia annulata</i>	<i>Protoperdinium brevipes</i>	
<i>Leptocylindrus danicus</i>	<i>Protoperdinium conicum</i>	
<i>Leptocylindrus minimus</i>	<i>Protoperdinium depressum</i>	
<i>Melosira moniliformis</i>	<i>Protoperdinium excentricum</i>	
<i>Odontella longircuris</i>	<i>Protoperdinium leonis</i>	
<i>Paralia sulcata</i>	<i>Protoperdinium oceanicum</i>	
<i>Rhizosolenia setigera</i>	<i>Protoperdinium</i> sp.	
<i>Skeletonema costatum</i>	<i>Protoperdinium steinii</i>	
<i>Stephanopyxis nipponica</i>	<i>Pyrophacus horologium</i>	
<i>Stephanopyxis palmeriana</i>	<i>Scrippsiella trochoidea</i>	
<i>Thalassiosira aestivalis</i>	diplopsalid dinoflagellate	
<i>Thalassiosira anguste-lineata</i>	gymnodiniod dinoflagellate	
<i>Thalassiosira eccentrica</i>	unidentified dinoflagellate (<25 um)	
<i>Thalassiosira nordenskioeldii</i>	unidentified dinoflagellate (>25 um)	
<i>Thalassiosira pacifica</i>		
<i>Thalassiosira punctigera</i>		
<i>Thalassiosira rotula</i>		
<i>Thalassiosira</i> sp.		
unidentified centric		

# 2008-2009 Dominant Taxa

## 1. Chain forming diatoms

*Chaetoceros* - many spp.

*Thalassiosira* - many spp.

*Skeletonema*

*Detonula*

*Rhizosolenia*

*Leptocylindrus*

*Dytilum*

*Eucampia*

*Thalassionema*

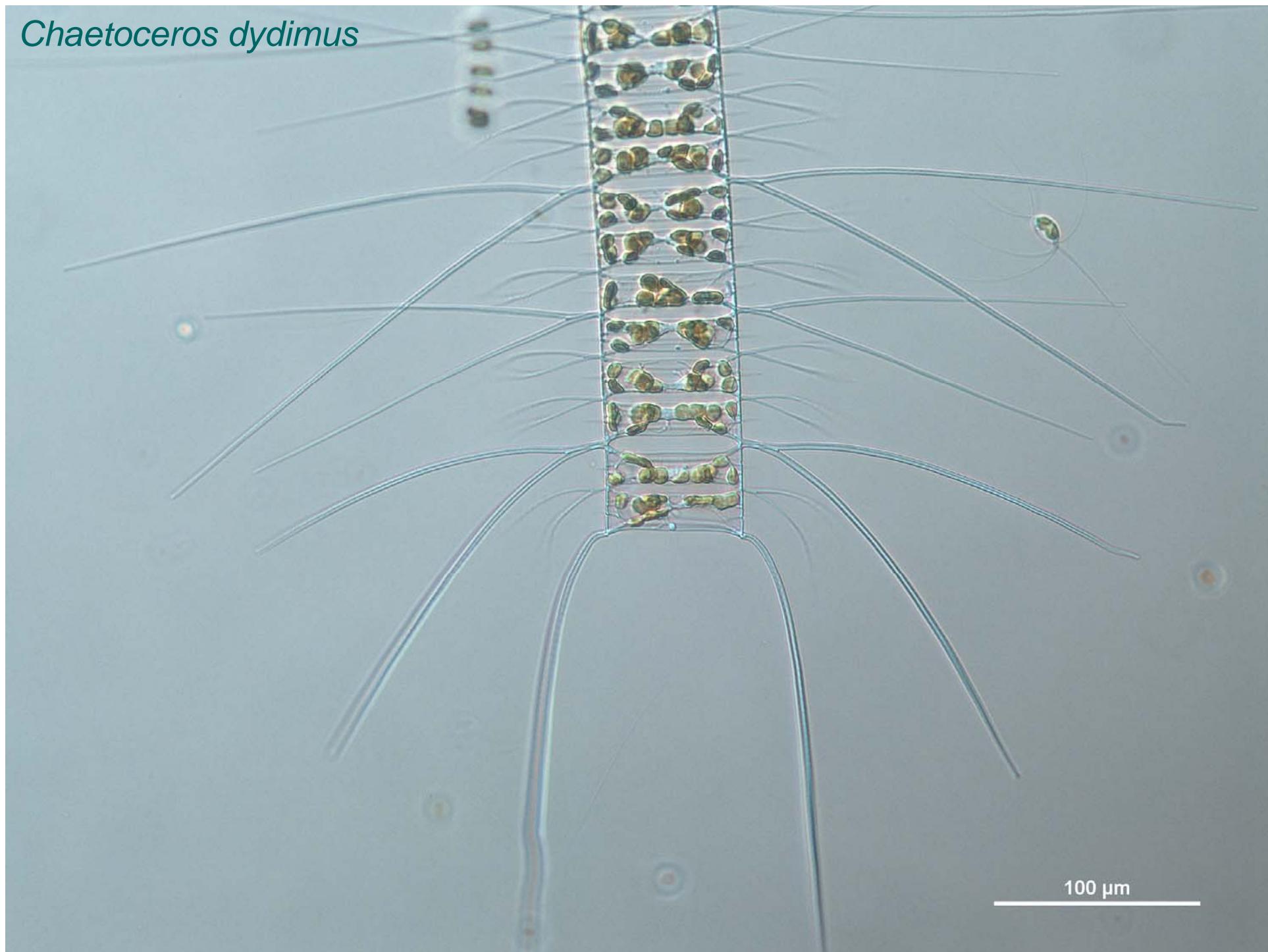
*Pseudo-nitzschia* (HAB)

*Chaetoceros debilis*



100 µm

*Chaetoceros dydimus*



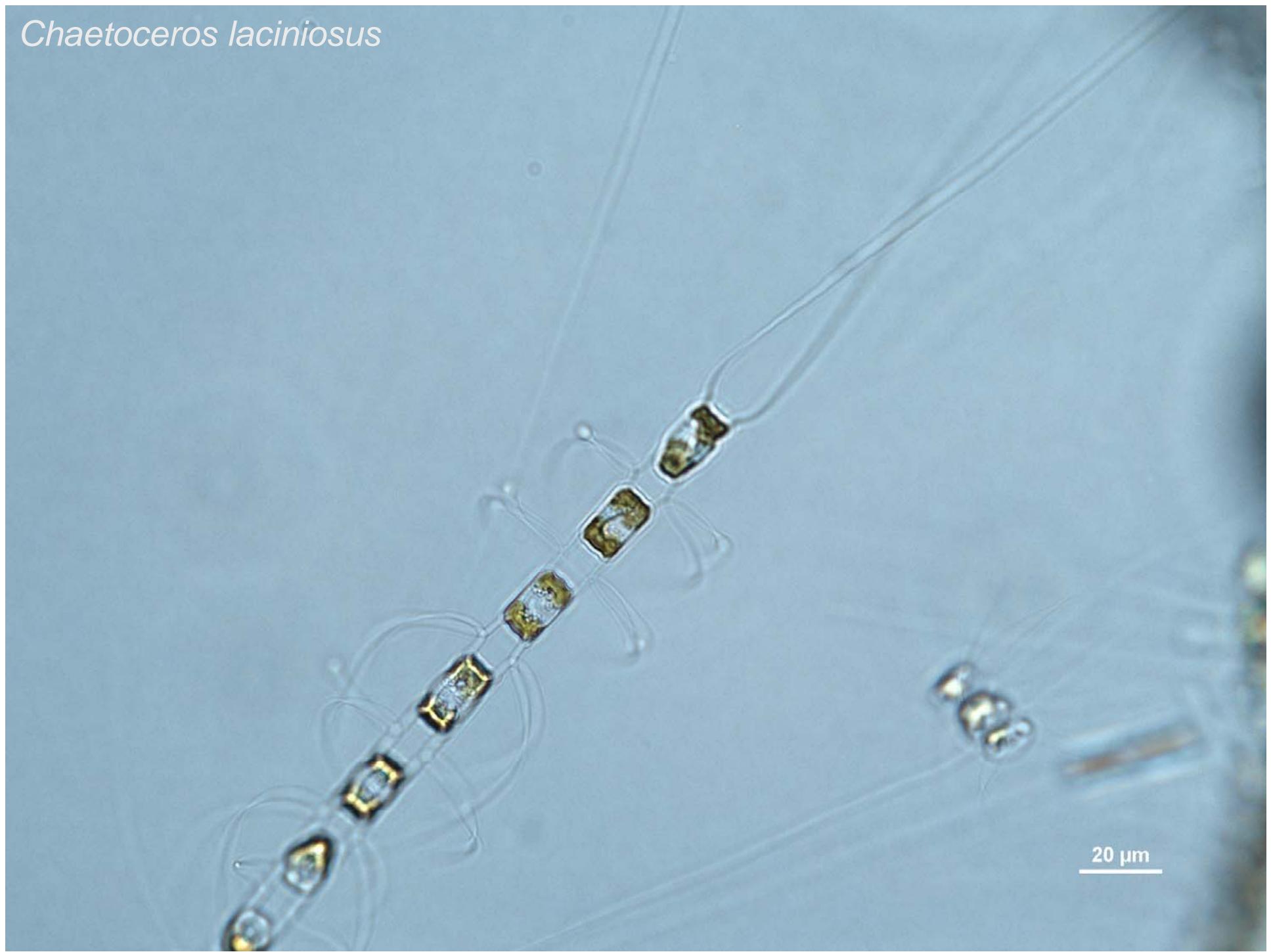
100 µm

*Chaetoceros diadema*



50  $\mu\text{m}$

*Chaetoceros laciniosus*



*Chaetoceros radicans*



50  $\mu\text{m}$

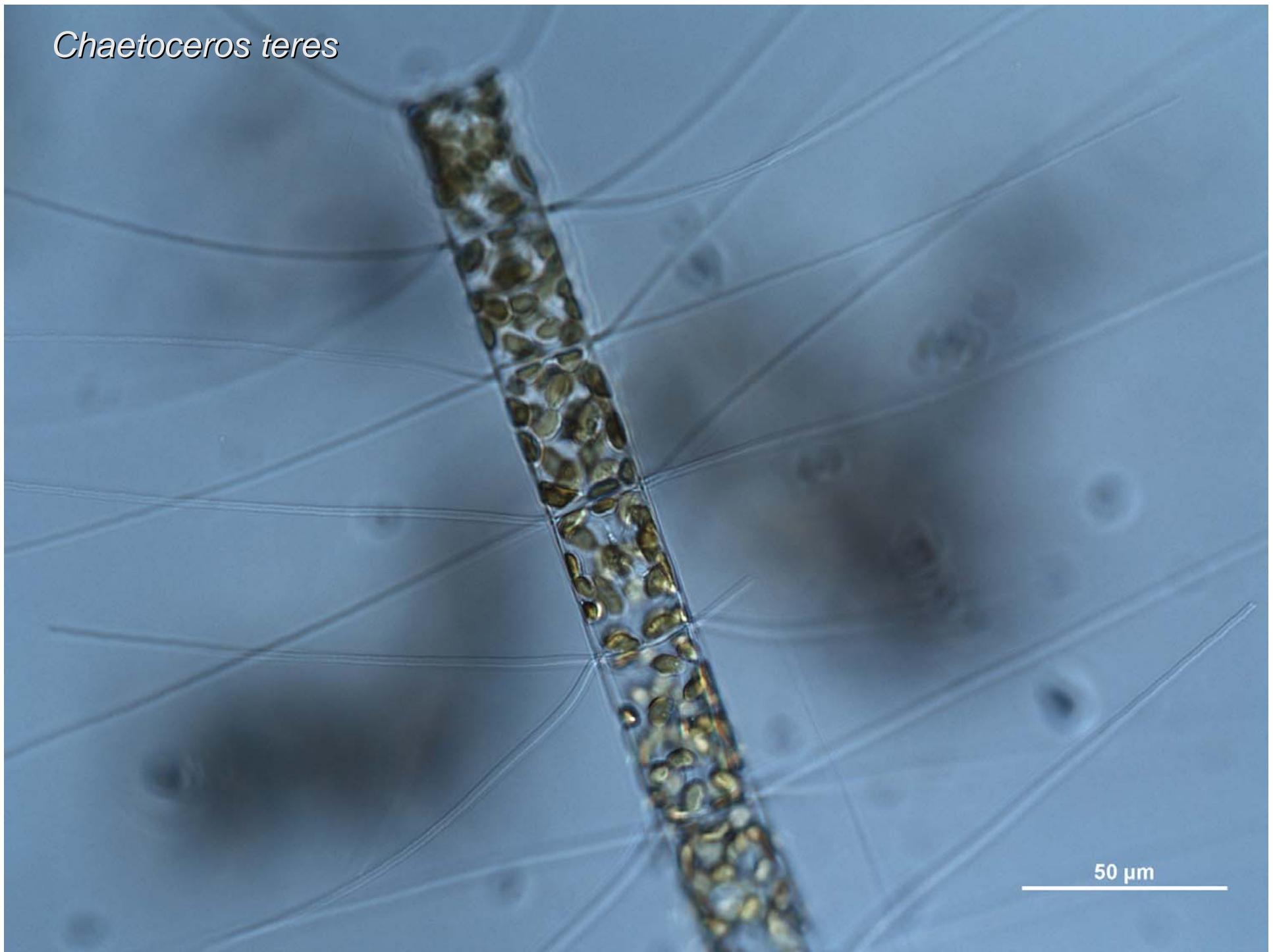
*Chaetoceros similis*



*Chaetoceros socialis*

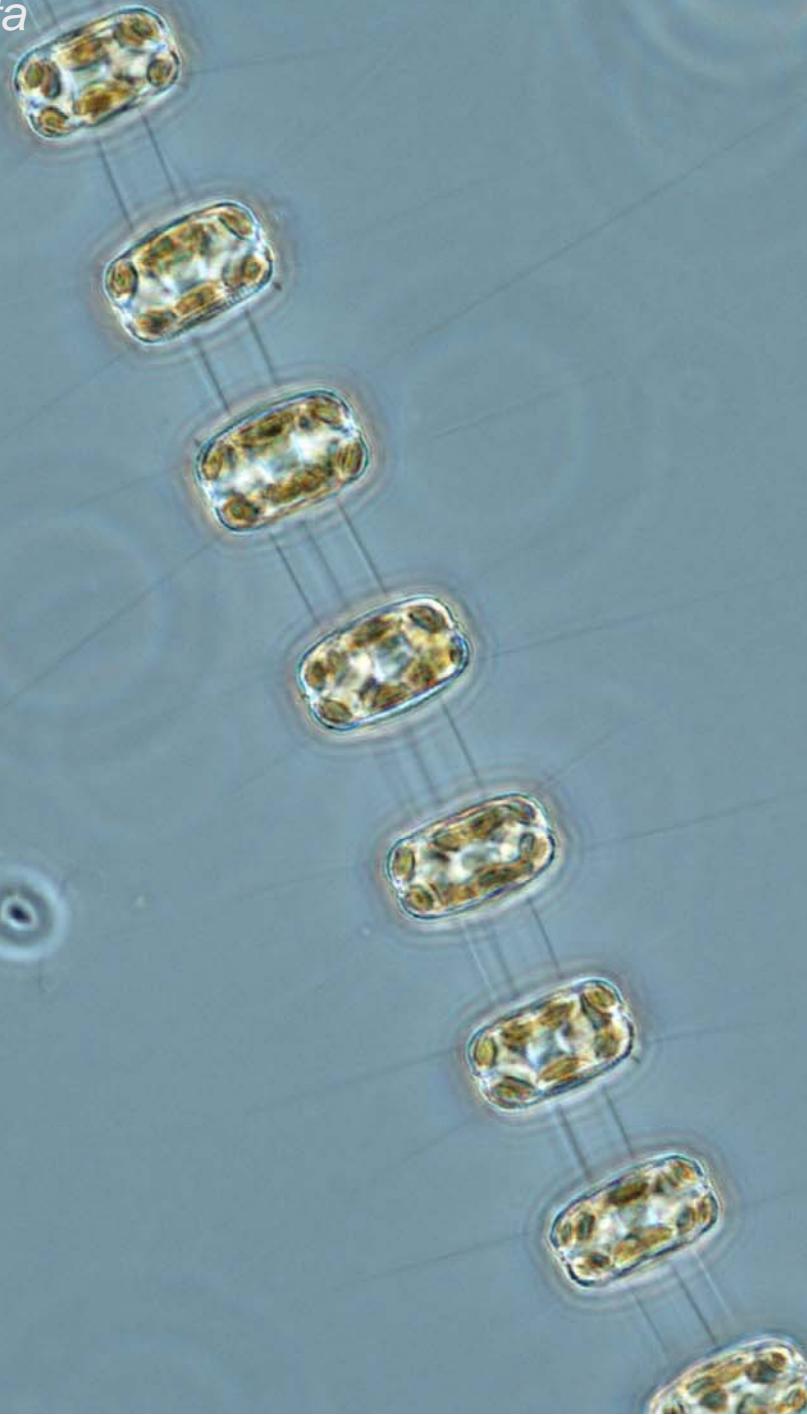


*Chaetoceros teres*



*Thalassiosira anguste-lineata*

50 µm



*Thalassiosira nordenskioeldii*



50 µm

*Skeletonema costatum*



*Ditylum brightwellii*



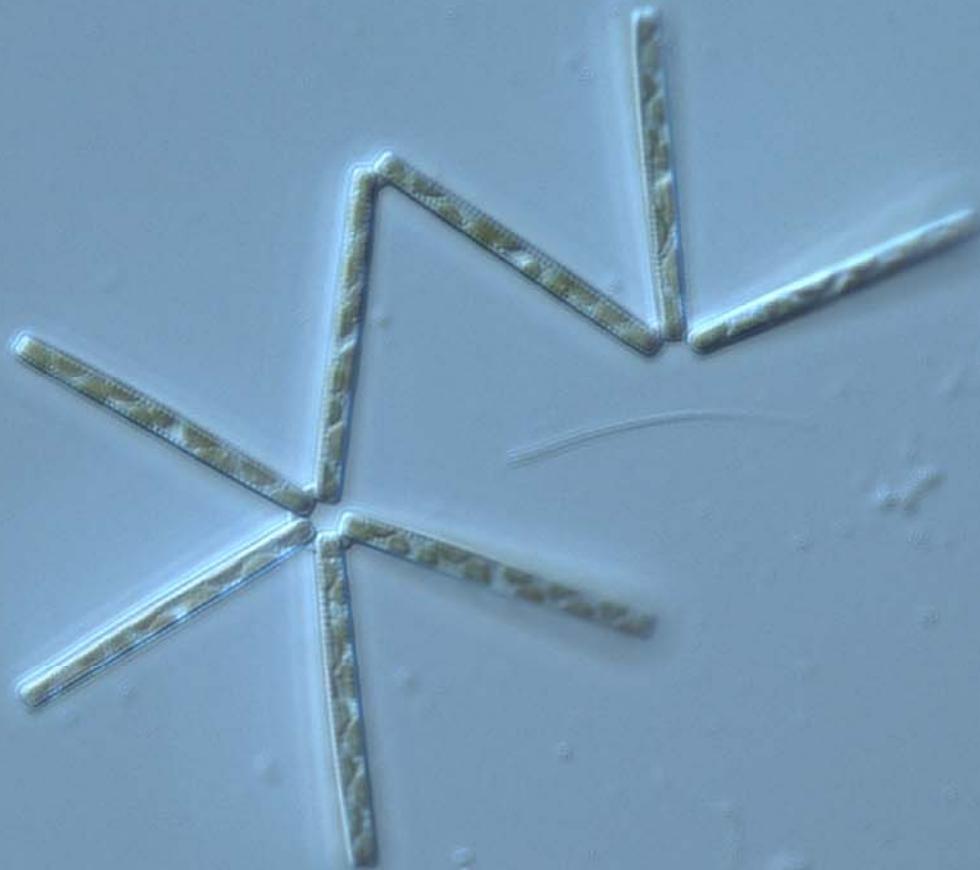
100  $\mu\text{m}$

*Rhizosolenia setigera*



100  $\mu\text{m}$

*Thalassionema nitzschiooides*



50  $\mu\text{m}$



*Pseudo-nitzschia* sp. (domoic acid)



50  $\mu\text{m}$

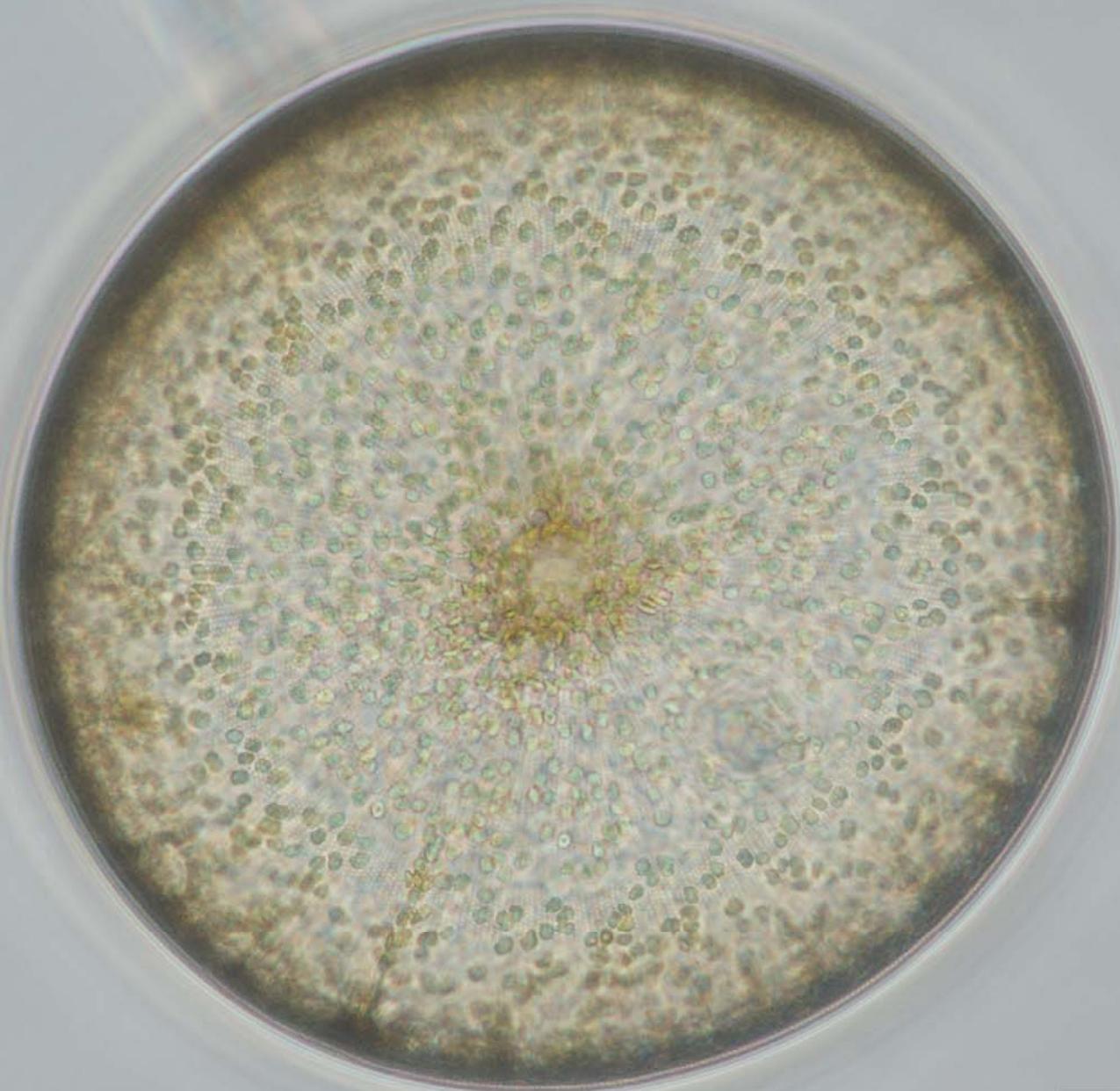
# 2008-2009 Dominant Taxa

## 2. Solitary diatoms

*Cylindrotheca*

*Coscinodiscus*

*Coscinodiscus wailessii*



100  $\mu\text{m}$

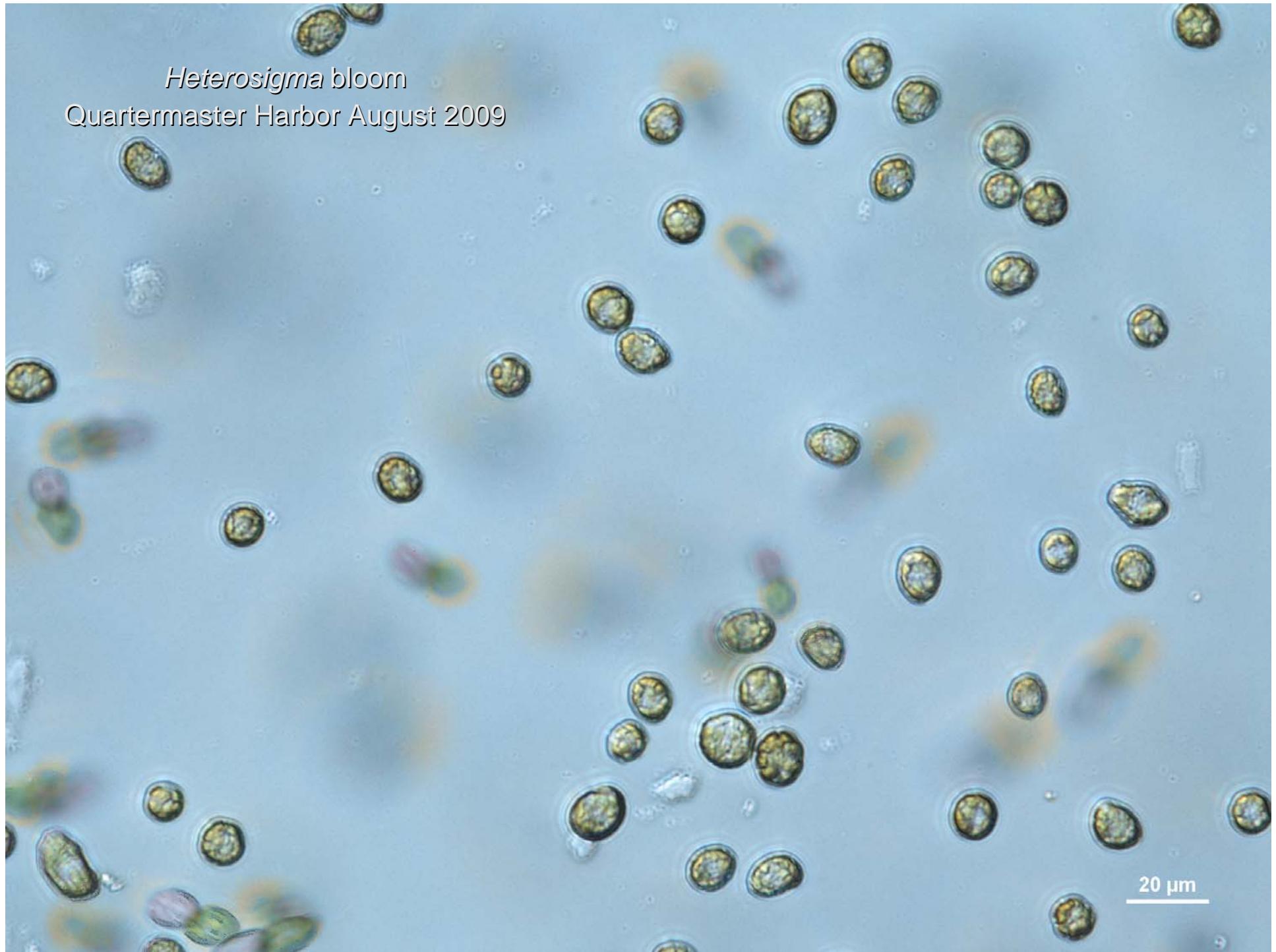
# 2008-2009 Dominant Taxa

## 3. *Heterosigma akashiwo*

Raphidophyte (small flagellate)

HAB

*Heterosigma* bloom  
Quartermaster Harbor August 2009



20  $\mu\text{m}$

# 2008-2009 Dominant Taxa

## 4. Dinoflagellates

*Protoperidinium* spp.

*Prorocentrum gracile*

*Ceratium fusus*

*Akashiwo sanguinea*

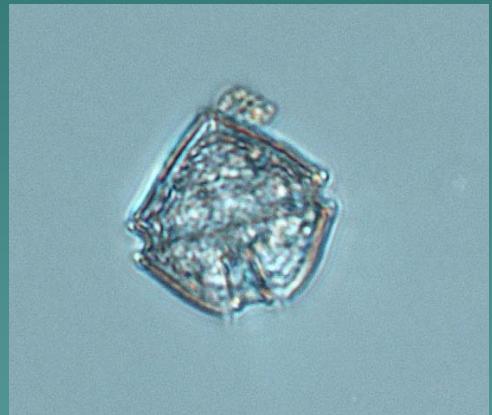
small unidentified spp.

*Alexandrium catenella* (PSP)

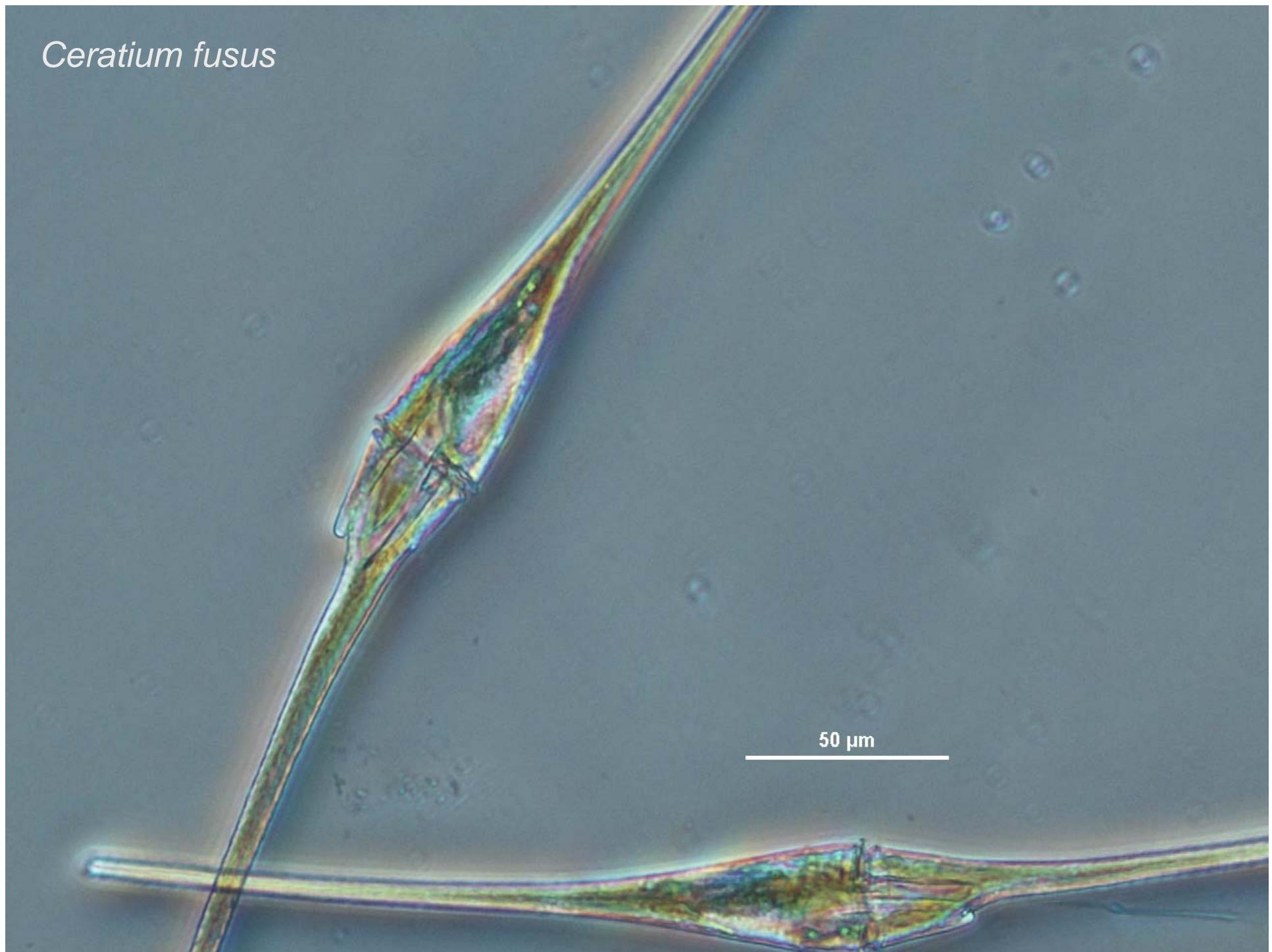
*Akashiwo sanguinea*



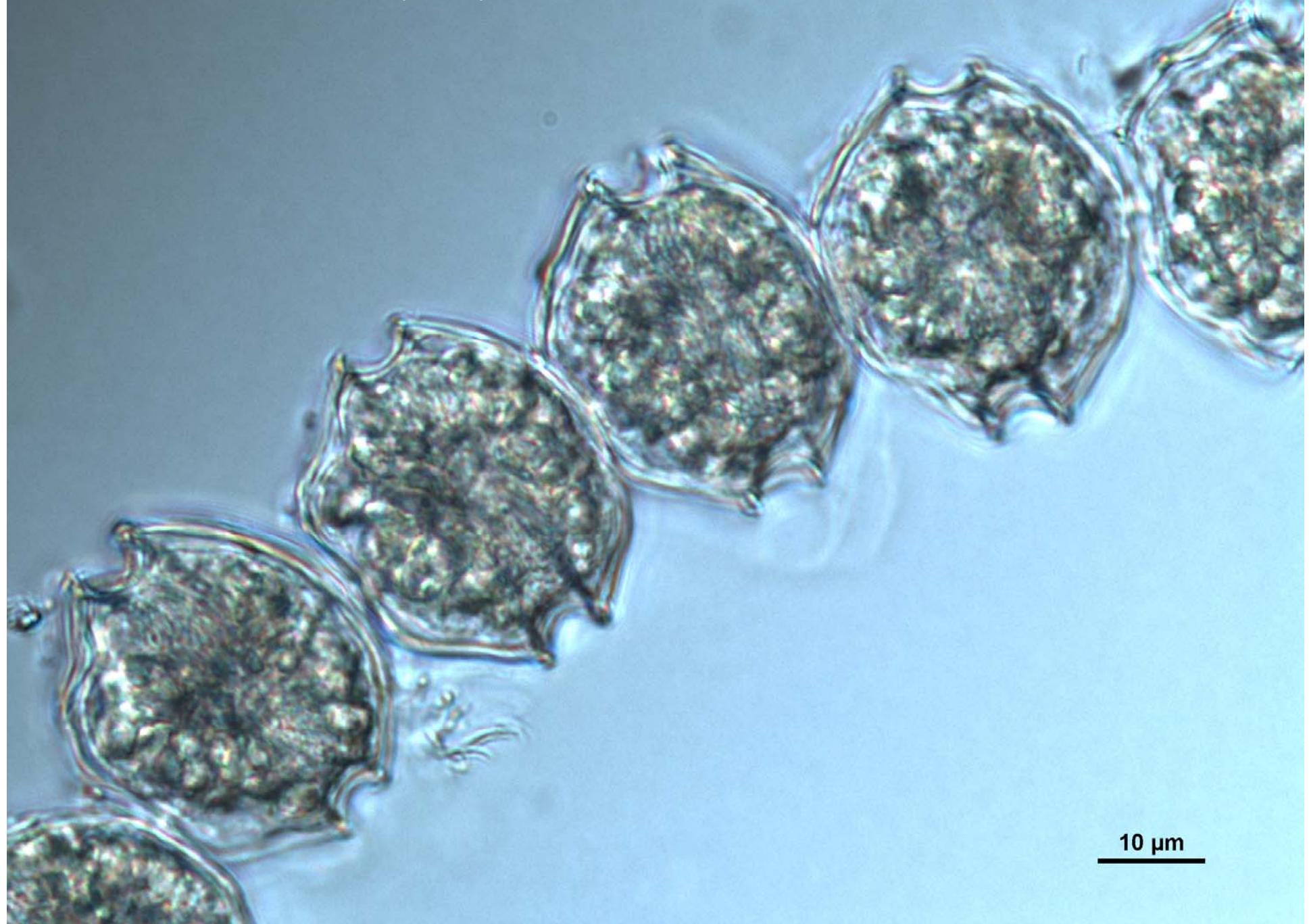
*Protoperidinium* spp.



*Ceratium fusus*



*Alexandrium catenella* (PSP)



# Other Taxa of Interest

or just  
more cool pictures

*Noctiluca* bloom



*Noctiluca scintillans*

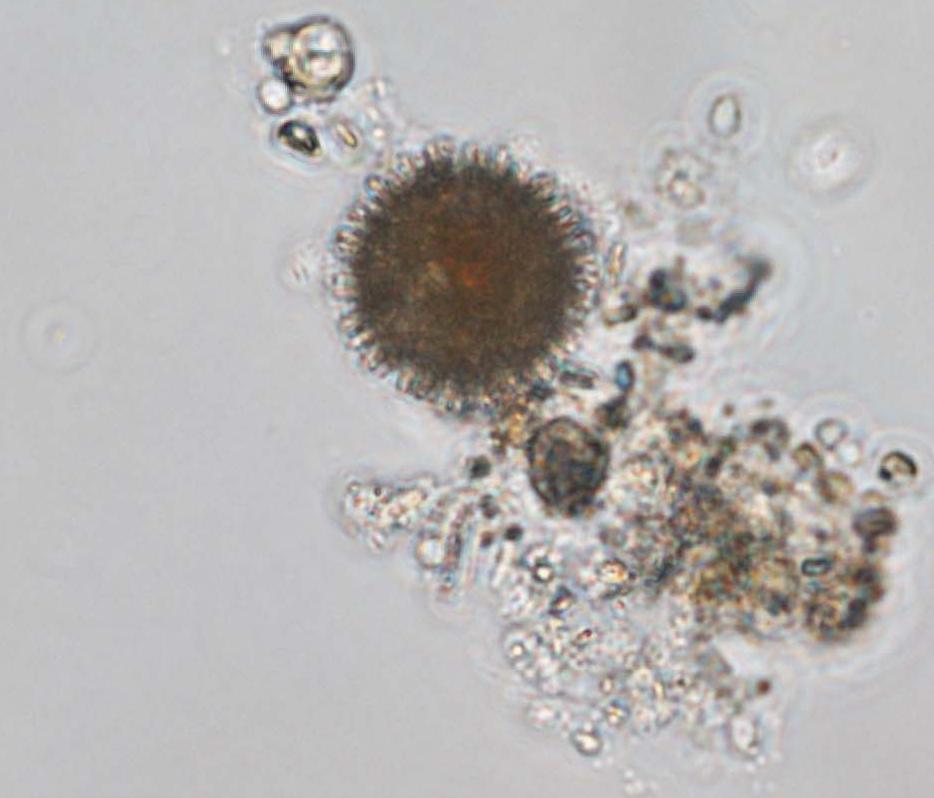


100  $\mu\text{m}$

*Dinophysis acuminata* (DSP)



Dinoflagellate cyst



*Gyrodinium* sp.



50  $\mu\text{m}$

*Gyrodinium spirale*



50 µm

# And what about the Zooplankton??



# Some Conclusions

- ◆ Challenges

- Sampling design: is it representative?

- Year to year variation

- “Correct” identification

- ◆ Use of technology?

- Flow-Cam for quantitative data

- ◆ Important effort!

- No other routine monitoring programs in the Sound

Acknowledgments: All involved staff + taxpayers